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# A Rapid and Sensitive HPLC Method for the Determination of Venlafaxine and O-Desmethylvenlafaxine in Human Plasma with UV Detection

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# A Rapid and Sensitive HPLC Method for the Determination of Venlafaxine and O-Desmethylvenlafaxine in Human Plasma with UV Detection

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## ABSTRACT

A rapid and sensitive high performance liquid chromatographic (HPLC) method was developed and validated for the analysis of venlafaxine (VEN) and O-desmethylvenlafaxine (ODV) in human plasma. The analytes were extracted from human plasma by using liquid–liquid extraction technique. Citalopram (CTP) was used as the internal standard. A Zorbax XDB C-18 column provided chromatographic separation of analytes followed by UV detection with wavelength at 229 nm. The linear

1297

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## 1298

#### Raut et al.

range for both analytes was 5 ng/mL to 1000 ng/mL with precision and accuracy of <10%. Total elution time was 12 min.

Key Words: Venlafaxine; ODV; Plasma; Citalopram.

# **INTRODUCTION**

Venlafaxine (VEN), 1-2-(dimethylamino)-1-(4-methoxyphenyl) ethylcyclohexanol hydrochloride, is a novel antidepressant. The pharmacologic action of VEN appears to involve the inhibition of neuronal uptake of catecholamines and serotonin. On the basis of clinical trials, this drug appears to lack many of the side effects associated with tricyclic antidepressants.

Venlafaxine has several metabolites, one of which is biologically active. About 1% of VEN is desmethylated to *N*-desmethylvenlafaxine, 1-(2-(methylamino)-1-(4-methoxyphenyl) ethyl) cyclohexanol; 16% becomes *N*,Odidesmethylvenlafaxine, 4-(2-(methylamino)-1-(1-hydroxycyclohexyl)ethyl) phenol; and 56% is metabolized to O-desmethylvenlafaxine (ODV), 4-(2dimethylamino)-1-(1-hydroxycyclohexyl) ethyl) phenol which, unlike the others, has an activity profile on monoamine transporters similar to VEN. Therefore, both VEN and ODV levels in plasma are important pharmacokinetic and pharmacodynamic parameters in assessing their efficacy.

Several methods have been reported in the literature for monitoring plasma levels of VEN and ODV.<sup>[1-15]</sup> The techniques used in these methods include mass spectrometric detection,<sup>[1,2]</sup> Capillary electrophoresis.<sup>[3–9]</sup> High performance liquid chromatography (HPLC) with fluorimetric detection,<sup>[10,11]</sup> HPLC with coulometric detection,<sup>[12]</sup> and HPLC with UV detection.<sup>[13–15]</sup>

This paper describes a simple and rapid HPLC method with UV detection for the determination of VEN and ODV in human plasma. This paper also describes the validation of the analytical procedure.

#### EXPERIMENTAL

## **Chemicals and Reagents**

The reference standards of VEN, and Citalopram HBr (CTP) were obtained from Wockhardt Research Centre (Aurangabad, India). O-Desmethylvenlafaxine was synthesized at Wockhardt Research Centre. High performance liquid chromatography quality water was prepared from an Elga water purification system. High performance liquid chromatography grade methanol, isopropyl alcohol, *n*-hexane, chloroform, and 1-octanesulphonic

## Venlafaxine and O-Demethylvenlafaxine by HPLC

1299

acid sodium salt were purchased from E. Merck Ltd. (Mumbai, India). Analytical reagent (AR) grade orthophosphoric acid, potassium dihydrogen phosphate, and disodium hydrogen phosphate were purchased from Qualigen fine chemicals (Mumbai, India). Drug free (blank) heparinized human plasma was obtained from Wockhardt Research Centre (Aurangabad, India).

## Instruments

The HPLC system consisted of a pump (Model Constametric 3500, Thermo Separation products, USA), an autosampler (Model AS 3000, Thermo Separation Products, USA), UV variable wavelength detector (Model UV 1000, Thermo Separation Products, USA), an interface (Model SP4510, Thermo Separation Products, USA), and a data station with PC 1000 software version 3.5.1. The analytical column used was Zorbax XDB-C18, 4.6 mm  $\times$  7.5 cm, 5 µm (Agilent Technologies, USA).

## **Chromatographic Conditions**

The mobile phase was a mixture of 62% water containing 0.05 M potassium dihydrogen phosphate, 28% methanol, and 10% isopropyl alcohol. It was filtered and degassed by passing through 0.45  $\mu$ m membrane filter paper (Millipore, USA). The flow-rate was 1.0 mL/min and the column temperature was 50°C. The total elution time was 12 min. The analytes were detected and quantified by UV detection at a wavelength of 229 nm.

#### **Preparation of Standard Solutions**

Stock solutions of VEN and internal standard CTP were prepared in water at free base concentration of  $1000 \,\mu\text{g/mL}$ . Secondary and working standard solutions were prepared by dilution with water. Stock solution of ODV was prepared by dissolving it in 10 mL of methanol and diluting with water at free base concentration of  $1000 \,\mu\text{g/mL}$ . Secondary and working standards were prepared by dilution with water.

#### **Extraction Procedure**

To a screw capped 15 mL glass test tube, 1 mL plasma and 50  $\mu$ L of the internal standard (0.8  $\mu$ g/mL) were added. The mixture was shaken on a

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## 1300

#### Raut et al.

vortex mixer for 30 sec. The pH of plasma was made alkaline by adding 0.2 mL of 0.5 M disodium hydrogen phosphate and 5 mL mixture of chloroform and *n*-hexane (70:30) was added. The mixture was shaken on a vortex mixer for 2 min and centrifuged for 10 min at 3000 rpm. About 4 mL of the organic layer was transferred to another 15 mL glass stoppered centrifuge tube containing 0.4 mL orthophosphoric acid (0.01% v/v). The mixture was shaken on a vortex mixer for 2 min and centrifuged for 10 min at 3000 rpm. The aqueous layer containing the drug was evaporated under a stream of nitrogen for 10 min at ambient temperature to evaporate any dissolved organic solvents. The residual aqueous phase was then transferred to an injection vial and 100  $\mu$ L of the extract was injected into the chromatographic system.

# Specificity and Linearity

The specificity of the method was evaluated with regards to interference due to the presence of endogenous substances in the extracted plasma. For the same, plasma pouches were randomly chosen for experimentation. Six different plasma samples were analysed. To establish the range of linearity for VEN and ODV, the concentrations 5, 10, 25, 50, 100, 200, 300, 500, 750, and 1000 ng/mL of VEN and ODV in plasma were used.

## Limit of Quantification

Limit of quantification was defined as the lowest concentration at which the precision expressed by relative standard deviation (RSD) is less than 20% and accuracy expressed by relative difference of the measured and true value was also less than 20%. To determine the limit of quantification, the plasma solutions containing 1, 2, 3, 4, 5, 6, 10, 25, 50, 100, 200, 300, 500, 750, and 1000 ng/mL of VEN and ODV in plasma were prepared and extracted as per the procedure and injected into the chromatographic system. The values were back calculated from the standard curve.

### Recovery

The recoveries of VEN, ODV, and CTP in the extraction procedure were determined by comparing the peak areas obtained from an extracted sample spiked with known amounts of VEN, ODV, and CTP with those obtained from the pure compounds of the same concentrations in the solutions. The

#### Venlafaxine and O-Demethylvenlafaxine by HPLC

1301

recoveries of VEN and ODV were determined at the concentrations of 10, 100, and 750 ng/mL. The recovery of CTP was determined at 400 ng/mL.

## **Precision and Accuracy**

To assess the accuracy and precision of the method, intra-day and interday (day 1, 2, 3, 4, and 5) measurements of VEN and ODV were completed and the relative error (% RE) and relative standard deviation (% RSD) for replicate samples (n = 5 for intra-day, n = 25 for inter-day) at concentration 5, 10, 100, and 750 ng/mL was determined. Both intra-day and inter-day samples were back calculated from calibration curves.

Accuracy expressed as % RE = [(mean/theoretical)-1] \* 100. Precision expressed as % RSD = (standard deviation/mean) \* 100.

### Data Calculation/Data Acceptability Criteria

Data calculation was accomplished with an online computer system. Calculation of drug concentration in unknown samples was based on a weighted  $(1/x^2)$  least squares regression of plasma calibrator concentrations against the peak area ratios. The peak area ratios were obtained by dividing the peak area of VEN and ODV by the peak area of the internal standard. Each calibration curve must have a coefficient of determination of at least 0.97. Both the relative error and the relative standard deviation of each level of the linearity curves and quality control samples must be <15% of the theoretical value except for those at the limit of quantification, which must be <20%.

#### RESULTS

### Chromatography and Specificity

High performance liquid chromatographic profiles for blank human plasma and plasma spiked with VEN, ODV, and CTP are shown in Fig. 1(A) and (B), respectively. The peaks of VEN, ODV, and CTP were well resolved, as shown in Fig. 1(B). No endogenous interfering peaks were found in blank plasma at their retention times [Fig. 1(A)], indicating a good selectivity for the chromatographic method.



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Raut et al.

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### Limit of Quantification

It was found that below 5 ng/mL, the back calculated values failed to meet the acceptance criteria. Hence, 5 ng/mL level was extracted five times and injected. It was found that relative standard deviation was 8.3% and the relative error was 5.0% at this concentration. So 5 ng/mL was established as the lowest limit of quantification.

## Standard Curve Characteristics, Precision, and Accuracy

The calibration curves were linear in the studied range. The slope, intercept, and correlation coefficient (mean  $\pm$  standard deviation) of standard curves (n = 5) for VEN was  $0.00322 \pm 0.00013$ ,  $0.00255 \pm 0.00338$ , and  $0.99938 \pm 0.00033$ , respectively. Similarly, for ODV, the slope, intercept, and correlation coefficient was  $0.00250 \pm 0.00015$ ,  $0.00614 \pm 0.00196$ , and  $0.99916 \pm 0.00062$ , respectively.

The back calculated concentration values for each level of VEN and ODV were obtained from each calibration curve used during method validation. The obtained mean (n = 5) back calculated values, as well as, the precision and accuracy estimations for each concentration level are summarized in Tables 1 and 2. The RSD and RE for VEN were 6.3% and -7.46%, respectively, and for ODV were 7.44% and -9.73%, respectively. The precision and accuracy

| Target value (ng/mL) | Mean $(ng/mL, n=5)$ | SD    | RE (%) | RSD (%) |
|----------------------|---------------------|-------|--------|---------|
| 5                    | 5.03                | 0.10  | 0.52   | 2.08    |
| 10                   | 9.97                | 0.48  | -0.32  | 4.78    |
| 25                   | 23.26               | 1.47  | -7.46  | 6.30    |
| 50                   | 53.88               | 1.23  | 7.19   | 2.28    |
| 100                  | 103.59              | 2.69  | 3.47   | 2.60    |
| 200                  | 198.23              | 5.00  | -0.89  | 2.52    |
| 300                  | 300.61              | 13.07 | 0.20   | 4.35    |
| 500                  | 489.64              | 14.88 | -2.12  | 3.04    |
| 750                  | 753.49              | 30.55 | 0.46   | 4.05    |
| 1,000                | 974.56              | 17.73 | -2.61  | 1.82    |

Table 1. Standard curve characteristics for VEN.

*Note:* SD, standard deviation; RE, relative error; RSD, relative standard deviation.

## Venlafaxine and O-Demethylvenlafaxine by HPLC

## 1305

| Target value (ng/mL) | Mean $(ng/mL, n=5)$ | SD    | RE (%) | RSD (%) |
|----------------------|---------------------|-------|--------|---------|
| 5                    | 4.85                | 0.24  | -3.14  | 4.94    |
| 10                   | 9.95                | 0.74  | -0.50  | 7.44    |
| 25                   | 22.78               | 0.93  | -9.73  | 4.07    |
| 50                   | 53.30               | 1.77  | 6.19   | 3.31    |
| 100                  | 102.05              | 1.83  | 2.01   | 1.80    |
| 200                  | 207.24              | 1.10  | 3.49   | 0.53    |
| 300                  | 295.40              | 21.82 | -1.56  | 7.39    |
| 500                  | 480.39              | 10.37 | -4.08  | 2.16    |
| 750                  | 747.91              | 32.01 | -0.28  | 4.28    |
| 1,000                | 970.26              | 11.06 | -3.06  | 1.14    |

Table 2. Standard curve characteristics for ODV.

*Note:* SD, standard deviation; RE, relative error; RSD, relative standard deviation.

for determination of VEN and ODV was evaluated for replicate analysis of the quality control samples at four different concentrations within the standard curve range (Table 3). Intra-day precision and accuracy for VEN and ODV was found to be less than 12% and inter-day precision and accuracy was less than 8%.

# **Stability Study**

## Freeze Thaw Stability

Freeze thaw stability was carried out at three different concentrations (10, 100, and 750 ng/mL). The plasma samples were stored at  $-40^{\circ}$ C and subjected to three freeze-thaw cycles. The results are represented in ng/mL from initial to after third cycle as (mean ± standard deviation). For concentrations of 10 ng/mL, 100 ng/mL, and 750 ng/mL of VEN, the results found were  $8.9 \pm 0.39$ ,  $96.34 \pm 2.02$ , and  $732.56 \pm 19.58$ . For concentrations of 10 ng/mL, 100 ng/mL, and 750 ng/mL of ODV, the results found were  $9.45 \pm 0.34$ ,  $93.05 \pm 4.71$ , and  $653.59 \pm 14.01$ . The concentrations found were well within the acceptable limit  $\pm 15\%$  of the nominal concentration, indicating no significant substance loss during repeated thawing and freezing.

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Raut et al.

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1306

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|       |       | Table 3. | Precision a | ind accuracy fi    | or VENL and | I ODV. |        |        |
|-------|-------|----------|-------------|--------------------|-------------|--------|--------|--------|
|       |       |          | Target cc   | oncentration (r    | ıg/mL)      |        |        |        |
|       |       | VE       | SN ST       |                    |             | 0      | DV     |        |
|       | 5     | 10       | 100         | 750                | 5           | 10     | 100    | 750    |
|       |       |          |             | Day 1 <sup>a</sup> |             |        |        |        |
| Mean  | 5.21  | 9.49     | 101.08      | 765.52             | 4.92        | 9.43   | 102.16 | 728.23 |
| SD    | 0.21  | 0.59     | 2.94        | 96.6               | 0.55        | 0.89   | 6.20   | 45.49  |
| % RE  | 4.29  | -5.14    | 1.08        | 2.07               | -1.65       | -5.70  | 2.16   | -2.90  |
| % RSD | 4.02  | 6.19     | 2.91        | 1.3                | 11.14       | 9.48   | 6.07   | 6.25   |
|       |       |          |             | Day 2 <sup>a</sup> |             |        |        |        |
| Mean  | 4.84  | 9.42     | 95.61       | 760.72             | 4.84        | 9.27   | 99.04  | 734.05 |
| SD    | 0.24  | 0.47     | 3.96        | 56.40              | 0.26        | 0.31   | 8.20   | 70.14  |
| % RE  | -3.15 | -5.81    | -4.39       | 1.43               | -3.11       | -7.30  | -0.96  | -2.13  |
| % RSD | 4.90  | 4.94     | 4.15        | 7.41               | 5.33        | 3.32   | 8.28   | 9.55   |
|       |       |          |             | Day 3 <sup>a</sup> |             |        |        |        |
| Mean  | 5.55  | 10.45    | 105.27      | 769.78             | 5.06        | 9.80   | 102.96 | 772.36 |
| SD    | 0.10  | 0.67     | 3.00        | 20.76              | 0.14        | 0.66   | 4.86   | 40.80  |
|       |       |          |             |                    |             |        |        |        |

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| % RE<br>% RSD                                     | -11.10<br>1.73            | -4.47<br>6.41 | -5.27<br>2.85 | 2.64<br>2.70                 | 1.14<br>2.71 | -2.04<br>6.73 | 2.96<br>4.72 | 2.98<br>5.28 |
|---|---------------------------|---------------|---------------|------------------------------|--------------|---------------|--------------|--------------|
| Mean  | د<br>13                   | 9 60          | 99 73         | Day 4 <sup>a</sup><br>768 38 | 4 90         | 0 84          | 100.21       | 773 88       |
| SD  | 0.31                      | 0.73          | 9.35          | 59.77                        | 0.31         | 0.54          | 10.50        | 35.53        |
| % RE  | 2.66                      | -3.96         | -0.27         | 2.45                         | -2.10        | -1.57         | 0.21         | 3.18         |
| % RSD   | 5.97                      | 7.57          | 9.37          | 7.78                         | 6.23         | 5.52          | 10.48        | 4.59         |
|   |                           |               |               | Day 5 <sup>a</sup>           |              |               |              |              |
| Mean  | 4.76                      | 10.04         | 99.24         | 711.57                       | 4.76         | 9.22          | 98.78        | 745.30       |
| SD  | 0.16                      | 1.00          | 6.92          | 10.37                        | 0.28         | 1.01          | 6.05         | 8.12         |
| % RE  | -4.77                     | 0.44          | -0.76         | -5.12                        | -4.77        | -7.81         | -1.22        | -0.63        |
| % RSD   | 3.45                      | 10.00         | 6.98          | 1.46                         | 5.95         | 10.94         | 6.13         | 1.09         |
|   |                           |               |               | Overall <sup>b</sup>         |              |               |              |              |
| Mean  | 5.10                      | 9.80          | 100.19        | 755.19                       | 4.90         | 9.51          | 100.63       | 750.76       |
| SD  | 0.35                      | 0.76          | 6.18          | 41.68                        | 0.32         | 0.71          | 6.99         | 45.19        |
| % RE  | 2.02                      | -2.00         | 0.19          | 0.69                         | -2.10        | -4.88         | 0.63         | 0.10         |
| % RSD   | 6.85                      | 7.79          | 6.17          | 5.52                         | 6.56         | 7.52          | 6.95         | 6.02         |
| $a^{\rm a}n = 5$ deter.<br>$b^{\rm b}n = 25$ dete | minations.<br>rminations. |               |               |                              |              |               |              |              |

Venlafaxine and O-Demethylvenlafaxine by HPLC

### 1308

#### Raut et al.

Processed Sample Stability

Processed sample stability was carried out at three different concentrations (10, 100, and 750 ng/mL). The samples were kept in an autosampler at ambient temperature and analysed at regular intervals up to 72 h. The results are presented in Tables 4 and 5. The processed samples were found to be stable at ambient temperature for 72 h.

#### Recovery

The mean percent recovery of VEN, ODV, and CTP from plasma, following the extraction procedure, was found to be 99.69%, 87.35%, and 73.02%, respectively.

## DISCUSSION

Several methods have been reported in the literature for monitoring plasma levels of VEN and ODV.<sup>[1-15]</sup> The techniques used in these methods include mass spectrometric detection,<sup>[1,2]</sup> Capillary electrophoresis,<sup>[3-9]</sup> HPLC with fluorimetric detection,<sup>[10,11]</sup> HPLC with coulometric detection,<sup>[12]</sup> and HPLC with UV detection.<sup>[13-15]</sup> Mass spectrometry and capillary electrophoresis are expensive techniques and would not be available in every laboratory. Fluorimetric and coulometric detectors are specific detectors. Considering the availability of these instruments and the cost involved, we have developed a new method, which utilises a UV detector which is a universal detector.

Also, out of the published methods, in the method utilising fluorimetric detection,<sup>[10]</sup> recovery of VEN, ODV, and IS was 100, 68, and 53%, respectively. The extraction procedure was tedious and time consuming (overnight freezing of the samples at  $-20^{\circ}$ C required).

Another method<sup>[12]</sup> utilising coulometric detection quoted the linearity range as 0–200 ng/mL. The reported  $C_{\text{max}}$  for 75 mg immediate release tablets were 225 ng/mL for VEN and 290 for ODV.<sup>[16]</sup> If the plasma levels exceed 200 ng/mL, the samples have to be diluted and reanalysed. Considering this, we have established the linearity range for VEN and ODV as 5–1000 ng/mL.

In the three previously published HPLC methods with UV detection, the first method<sup>[13]</sup> has quoted the limit of quantification for VEN and ODV as 200 ng/mL. The second method<sup>[14]</sup> was used to determine VEN only and not ODV in human serum. The limit of quantification for VEN was 25 ng/mL.

The third method<sup>[15]</sup> has quoted the limit of quantification as 10 ng/mL. The procedure involves a total analysis time of 100 min for one sample, which includes shaking, centrifugation, and chromatographic runs. In comparison, in

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# Venlafaxine and O-Demethylvenlafaxine by HPLC

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|  |            | Tabl | e 4. Processed | sample stabi | lity for VEN. |      |          |      |
|--|------------|------|----------------|--------------|---------------|------|----------|------|
| 0<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2<br>2 | Initi      | al   | After 2        | 24 h         | After 4       | 48 h | After 7  | 72 h |
| sample conc. (ng/mL)   | Measured   | % RE | Measured       | % RE         | Measured      | % RE | Measured | % RE |
| 10   | 10.35      | 3.50 | 10.68          | 6.80         | 10.42         | 4.20 | 10.22    | 2.20 |
| 100  | 103.19     | 3.19 | 105.58         | 5.58         | 106.55        | 6.55 | 106.03   | 6.03 |
| 750  | 769.72     | 2.63 | 787.62         | 5.02         | 779.84        | 3.98 | 781.35   | 4.18 |
| Note: RE, relati   | ive error. |      |                |              |               |      |          |      |

1310

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|                         |           | Tal   | ble 5. Processe | d sample stabi | lity for ODV. |        |          |       |
|-------------------------|-----------|-------|-----------------|----------------|---------------|--------|----------|-------|
| Councilo acono          | Initi     | al    | After 2         | 24 h           | After 4       | 48 h   | After 7  | 72 h  |
| sample conc.<br>(ng/mL) | Measured  | % RE  | Measured        | % RE           | Measured      | % RE   | Measured | % RE  |
| 10                      | 9.19      | -8.10 | 8.96            | -10.40         | 8.93          | -10.70 | 9.69     | -3.1  |
| 100                     | 100.41    | 4.10  | 102.79          | 2.79           | 103.73        | 3.73   | 102.89   | 2.89  |
| 750                     | 738.59    | -1.52 | 746.69          | -0.44          | 742.66        | -0.98  | 741.51   | -1.13 |
| Note: RE, relati        | ve error. |       |                 |                |               |        |          |       |

Raut et al.

#### Venlafaxine and O-Demethylvenlafaxine by HPLC

1311

the newly developed method, only 46 min are required for analysing one sample. Also the limit of quantification established is 5 ng/mL. Thus, the current method reduces the analysis time of each sample by approximately 50% and is more sensitive than the published method.

In this study, we have developed a specific, accurate, and precise analytical method for the simultaneous determination of VEN and ODV in human plasma. The limit of quantification for VEN and ODV, each, has been established as 5 ng/mL. The total elution time was about 12 min. The newly developed method is more rapid and sensitive than previously published HPLC methods with UV detection. The inter-day precision expressed as %RSD for VEN and ODV was less than 8%. The inter-day accuracy expressed as %RE for VEN and ODV was between -2.10% and 2.02%. The intra-day precision expressed as %RSD for VEN and ODV was less than 11.5%. The intra-day accuracy expressed as %RE for VEN and ODV was between -11.10% and 11.14%.

Adjusting the solvent composition, mobile phase, pH, and column length optimised the presently described chromatographic method. In the present extraction procedure, recovery of VEN was approximately 100% over the nominal concentration of 10, 100, and 750 ng/mL. The recovery of ODV and CTP was approximately 87% and 73%, respectively. Increasing the composition of hexane in the extraction solvent resulted in an increase in the recovery of ODV, however, the recovery of VEN and CTP decreased. Trace amounts of organic solvents in the aqueous phase (0.01% v/v orthophosphoric acid) resulted in interfering peaks in close proximity of ODV. Evaporation of the aqueous phase for 10 min was, therefore, necessary to remove the traces of organic solvents.

The  $C_{\text{max}}$  values of VEN and ODV for 75 mg VEN immediate release tablets are 225 ng/mL and 290 ng/mL. The  $C_{\text{max}}$  values of VEN and ODV for 150 mg VEN extended release capsules are 150 ng/mL and 260 ng/mL.<sup>[16]</sup>

In the present method, we have established a linearity range of 5-1000 ng/mL. This linearity range covers the  $C_{\text{max}}$  values of the above mentioned strengths of VEN. Hence, this method can be applied for quantifying the levels of VEN and ODV in vivo, when either of the strengths is administered to the subjects.

#### **CONCLUSION**

A HPLC method for the determination of VEN and ODV in human plasma has been developed and validated. It has been shown to be rapid, accurate, precise, and sensitive. The statistical analysis of the precision and

#### Raut et al.

accuracy data demonstrates that the method is suitable for quantifying the two analytes during clinical trials and/or pharmacokinetic studies to a limit of quantification of 5 ng/mL.

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# REFERENCES

- 1. Cherkaoui, S.; Rudaz, S.; Varesio, E.; Veuthey, J.L. Online capillary electrophoresis–electrospary mass spectrometry for the stereoselective analysis of drugs and their metabolites. Electrophoresis **2001**, *22* (15), 3308–3315.
- Cherkaoui, S.; Rudaz, S.; Veuthey, J.L. Nonaqueous capillary electrophoresis-mass spectrometry for separation of venlafaxine and phase I metabolites. Electrophoresis 2001, 22 (3), 491–496.
- Desiderio, C.; Aturki, Z.; Fanali, S. Use of vancomycin silica stationary phase in packed capillary electrochromatography. I. Enantiomer separation of basic compounds. Electrophoresis 2001, 22 (3), 535–543.
- Fanali, S.; Rudaz, S.; Veuthey, J.L.; Desiderio, C. Use of vancomycin silica stationary phase in packed capillary electrochromatography. II. Enantiomer separation of venlafaxine and O-desmethylvenlafaxine in human plasma. J. Chromatogr. A. 2001, *919* (1), 195–203.
- Fanali, S.; Catarcini, P.; Quaglia, M.G. Use of vancomycin silica stationary phase in packed capillary electrochromatography. III. Enanatiomeric separation of basic compounds with the polar organic mobile phase. Electrophoresis 2002, 23 (3), 477–485.
- Rudaz, S.; Stella, C.; Balant-Gorgia, E.; Fanali, S.; Veuthey, J.L. Simultaneous stereoselective analysis of venlafaxine and O-desmethylvenlafaxine enantiomers in clinical samples by capillary electrophoresis using charged cyclodextrins. J. Pharm. Biomed. Anal. 2000, 23 (1), 107–115.
- Rudaz, S.; Veuthey, J.L.; Desiderio, C.; Fanali, S. Enantioseparation of venlafaxine and O-desmethylvenlafaxine by capillary electrophoresis with mixed cyclodextrins. Chromatografia **1999**, *50* (5–6), 369–372.

#### Venlafaxine and O-Demethylvenlafaxine by HPLC

#### 1313

- 8. Fanali, S.; Cotichini, V.; Porra, R. Analysis of venlafaxine by capillary zone electrophoresis. J. Capillary Electrophor. **1997**, *4* (1), 21–26.
- Labat, L.; Deveaux, M.; Dallet, P.; Dubost, J.P. Separation of new antidepressants and their metabolites by micellar electrokinetic capillary chromatography. J. Chromatogr. B. Analyt. Technol. Biomed. Life Sci. 2002, 773 (1), 17–23.
- Vu, R.L.; Helmeste, D.; Albers, L.; Reist, C. Rapid determination of venlafaxine and O-desmethylvenlafaxine in human plasma by highperformance liquid chromatography with fluorimetric detection. J. Chromatogr. B. **1997**, *703* (1–2), 195–201.
- 11. Li, H.D.; Ding, D.R.; Zhang, B.K.; Yuan, H.Y. Studies on determination of the new drug venlafaxine in human plasma by HPLC with fluorimetric detection **2001**.
- Clement, E.M.; Odontiadis, J.; Franklin, M. Simultaneous measurement of venlafaxine and its major metabolite, oxydesmethylvenlafaxine, in human plasma by high-performance liquid chromatography with coulometric detection and utilization of solid- phase extraction. J. Chromatogr. B. Biomed. Sci. Appl. **1998**, *705* (2), 303–308.
- Matoga, M.; Pehourcq, F.; Titier, K.; Dumora, F.; Jarry, C. Rapid high performance liquid chromatographic measurement of venlafaxine and O-desmethylvenlafaxine in human plasma. Application to management of acute intoxication. J. Chromatogr. B. Biomed. Sci. Appl. 2001, 760 (2), 213–218.
- Tournel, G.; Houdret, N.; Hedouin, V.; Deveau, M.; Gosset, D.; Lhermitte, M. High performance liquid chromatographic method to screen and quantitate seven selective serotonin reuptake inhibitors in human serum. J. Chromatogr. B. Biomed. Sci. Appl. 2001, 761 (2), 147–158.
- Hicks, D.R.; Wolaniuk, D.; Russell, A.; Cavanaugh, N.; Kraml, M. High performance liquid chromatographic method for the simultaneous determination of venlafaxine and O-desmethylvenlafaxine in biological fluids. Ther. Drug Monit. **1994**, *16* (1), 100–107.
- Physicians Desk Reference, 56th Ed.; Kelly, G.L., Murray, L., Sifton, D.W., Eds., Medical Economics Company, Inc.: Montvale, NJ, 2002; 3495–3504.

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